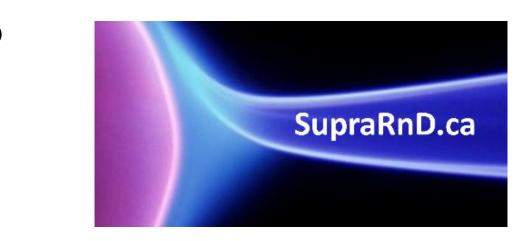
Quantifying Glycosidically-Bound Sensory Precursors in Smoke-Exposed Vitis vinifera Berries

Matthew Noestheden^{1,2}, Eric Dennis¹, Katelyn Theissen¹, Brandon Whitemore¹, Ben Tiet¹ & Wesley Zandberg¹

- ¹ University of British Columbia Okanagan, Kelowna, British Columbia, Canada
- ² Supra Research & Development, Kelowna, British Columbia, Canada





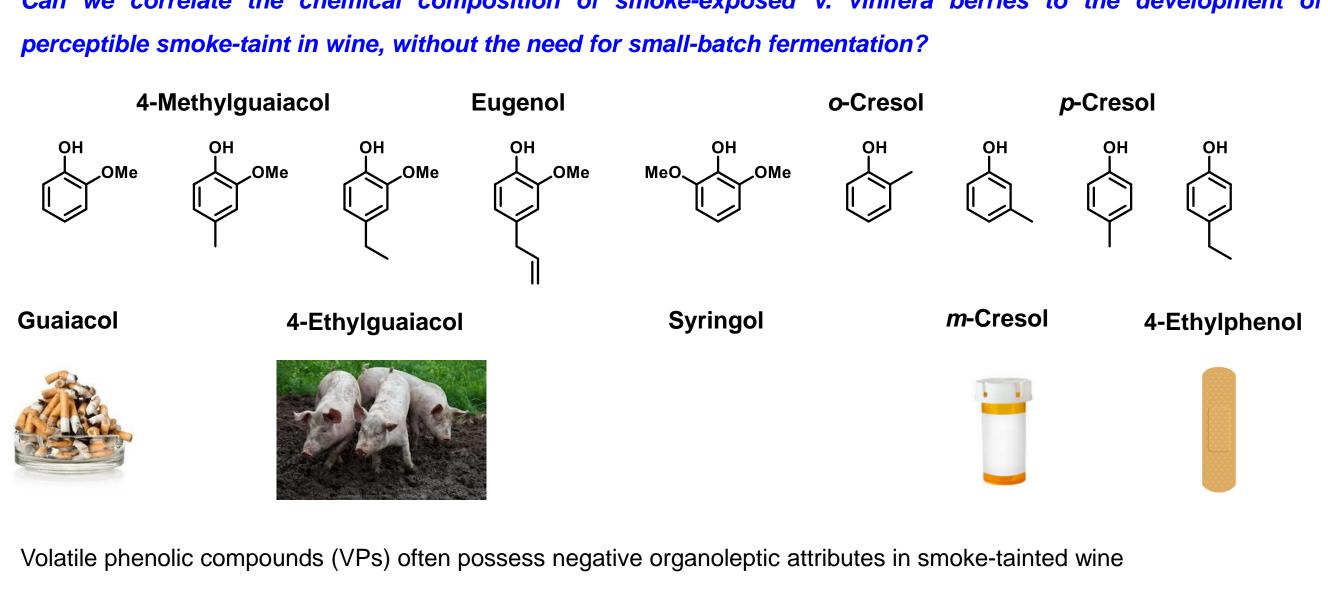
Abstract

Volatile phenolic compounds (e.g., guaiacol) and their sugar-bound glycosides are known to negatively influence the sensory attributes of wine made from smoke-exposed *Vitis vinifera* (*V. vinifera L*) berries and have been correlated with unpleasant 'smoky', 'ashy', 'burnt meat' and 'Band-Aid' aromas (*i.e.*, smoke-taint). Understanding this phenomenon is paramount, given that much of the North American wine grape crop is produced near forest fire-prone regions. To date, most volatile phenolic glycosides have been putatively assigned in the absence of direct empirical evidence. While these glycosides are not sensory-active, they are metabolised during fermentation to release volatile phenolic compounds that have distinct sensory descriptors. Therefore, phenolic glycosides constitute a 'sensory potential' that can negatively impact the flavour and aroma of wine following fermentation. It follows then, that the lack of definitive glycoside characterization is problematic since it: a) hinders efforts to develop remedial and preventative strategies; b) precludes absolute quantitative assessment of the total pool of volatile phenolic compounds; and c) confounds any correlation between volatile phenolics and their potential impact on wines made from smoke-exposed berries. We have developed an accurate analytical workflow capable of quantifying phenolic glycosides in *V. vinifera* berries. The systematic approach employed enabled the identification of several issues with existing methods that limit their quantitative accuracy. These deficiencies were critical since they impacted the generation of meaningful risk-assessment strategies based on total volatile phenol load when using smoke-exposed berries. The adaptability of our proposed strategy will enable its use on other glycosylated metabolites integral to wine quality (e.g., terpenoids or norisoprenoids).

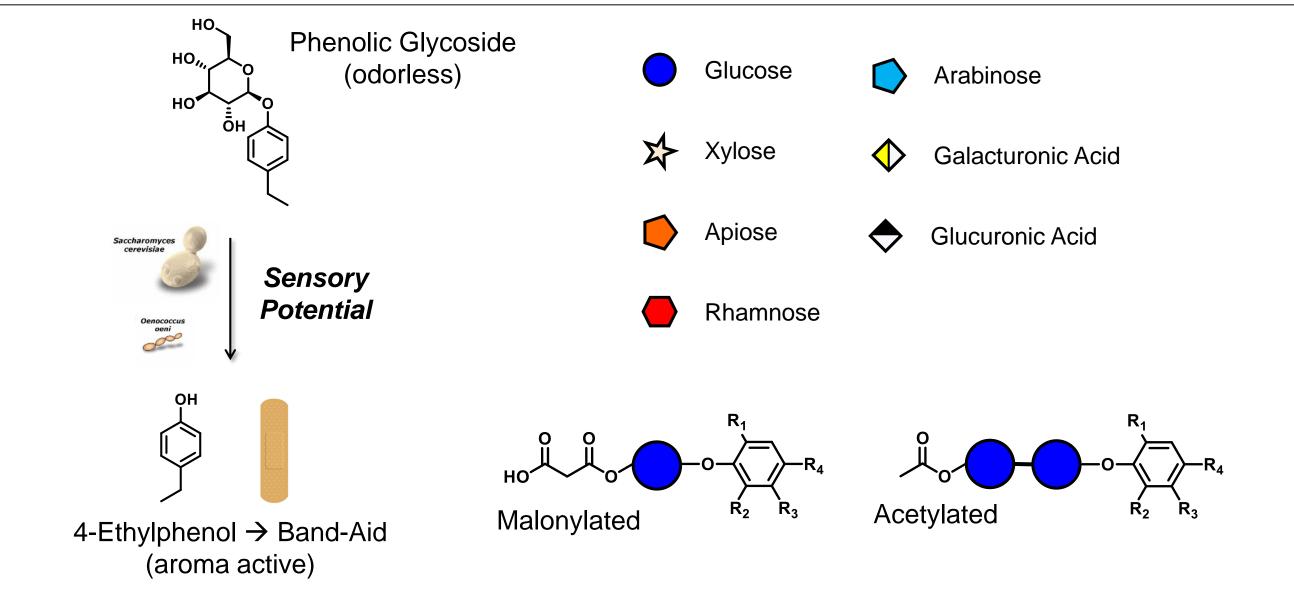
Background



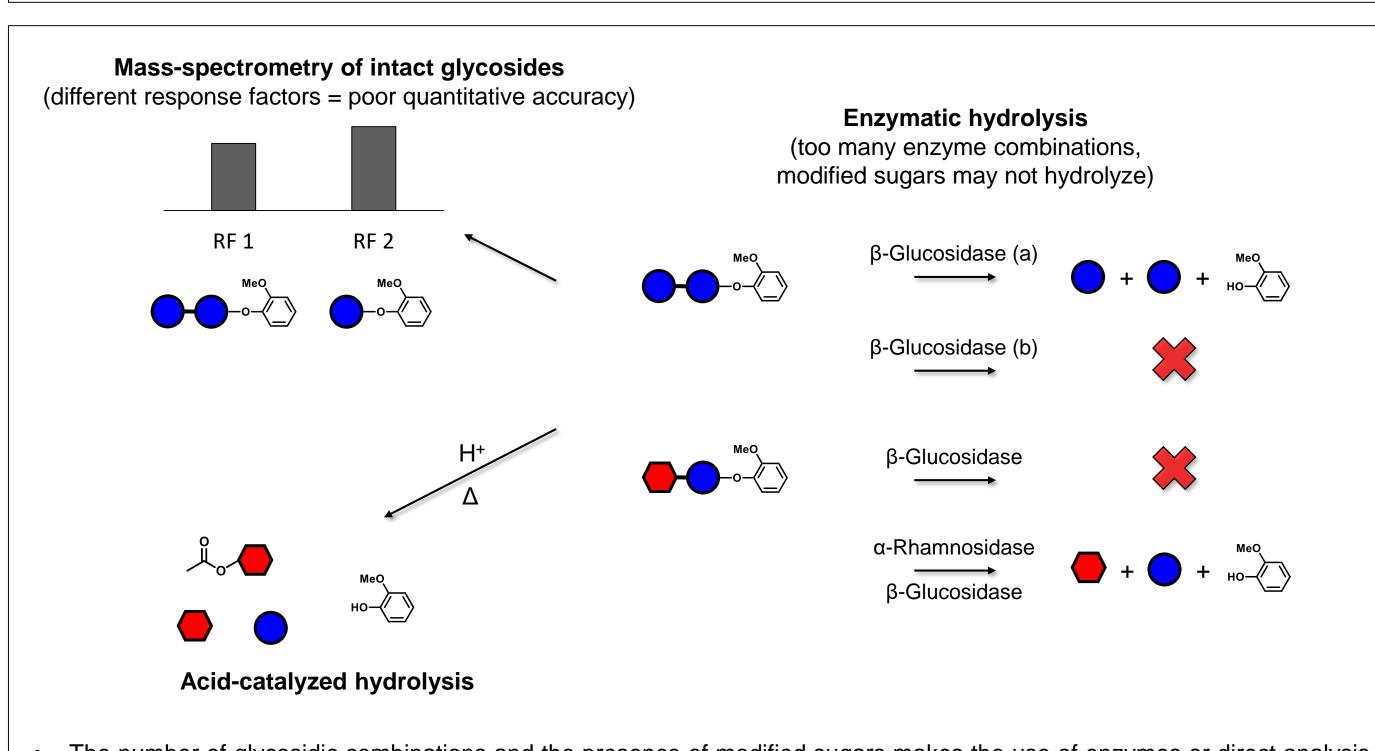
- Early prediction of crop issues is key to mitigating risk
- Can we correlate the chemical composition of smoke-exposed V. vinifera berries to the development of



- Volatile phenolic compounds (VPs) often possess negative organoleptic attributes in smoke-tainted wine
- VPs are only 50-80%^a predictive of perceptible smoke-taint



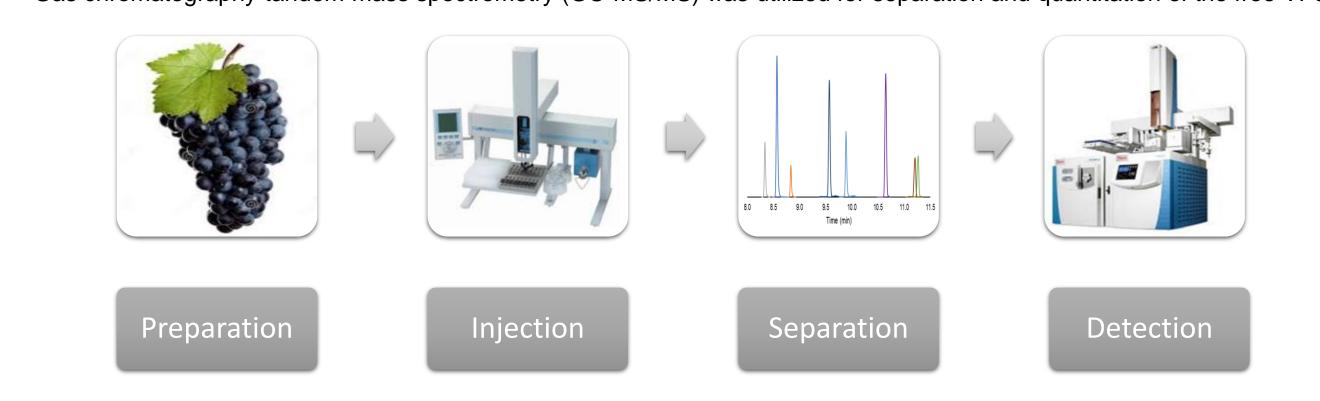
- 'Sensory potential' will determine the total VP load available for release during fermentation
- The glycosidically-bound forms are often present at 2-8x higher concentrations b than the free VPs
- The number of potential glycosides makes their direct detection analytically challenging



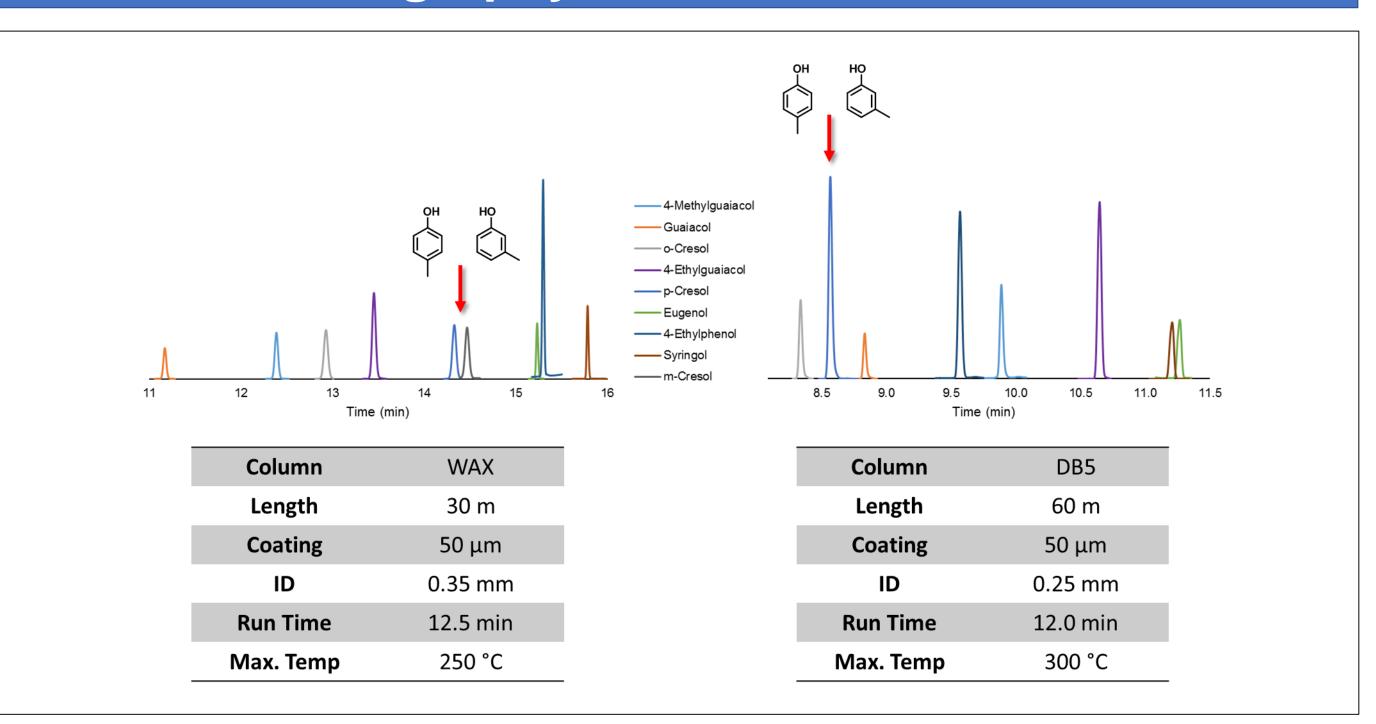
- The number of glycosidic combinations and the presence of modified sugars makes the use of enzymes or direct analysis via mass-spectrometry challenging, especially if quantitative accuracy is paramount
- Acid-catalyzed hydrolysis is a non-specific hydrolytic approach that should capture all of the potential glycosides and convert them to free VPs, where accurate quantitation is possible

Methods

• Gas chromatography-tandem mass spectrometry (GC-MS/MS) was utilized for separation and quantitation of the free VPs



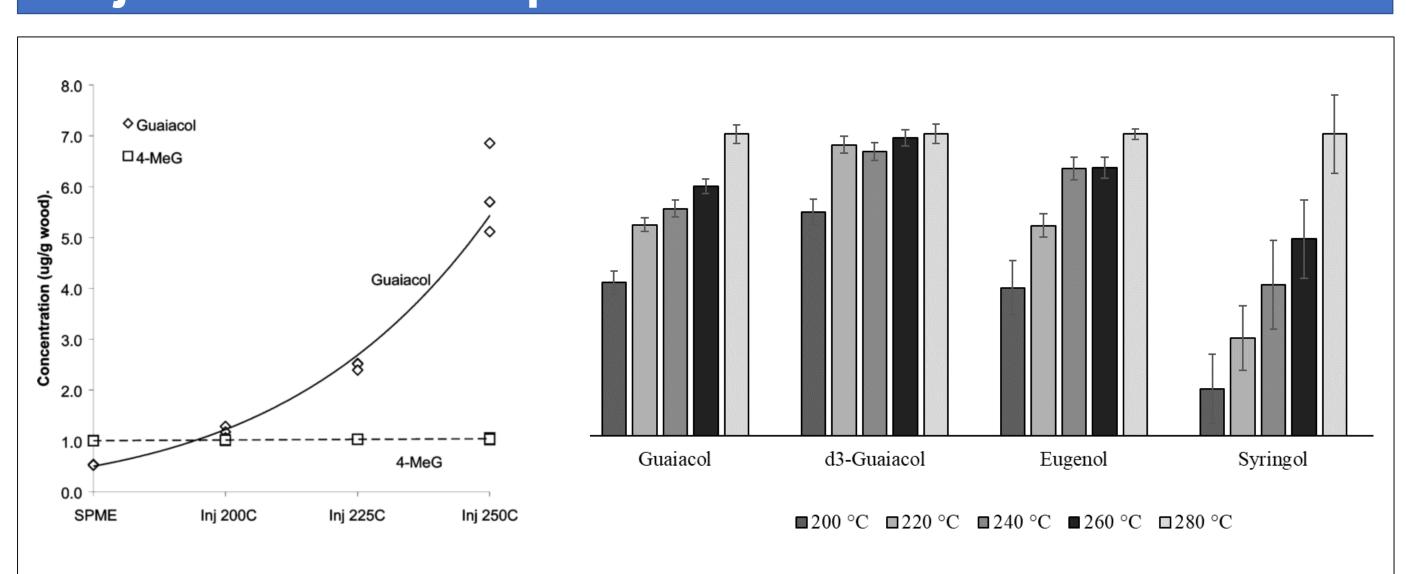
Gas Chromatography



Sample Preparation

Extraction Solvent	NaCl _{Sat}	Matrix	Inj. Temp. (°C)	 A variety of liquid-liquid extraction conditions have been published, but no
1:1 Hexane:EtOAc f	Υ	Berries	240	systematically compared
2:1 Pentane:Ether c	-	Wine	200	 Hypothesized differences in extraction
EtOAc	Υ	Berries	220	efficiency would impact quantitation of VPs
Hexane ^e	Υ	Berries	250	 Five extraction conditions were evaluated
Pentane d	-	Wine	200	(left) to identify optimal parameters
O.0 Country leguatacol A. Meetry leguatacol	Hexane:EtOAc	Ethyleuniacol .	Ether EtOAc	Hexane Pentane
1:1 hexane:EtOAc with satuMost compounds were reco	•			on efficiency for all compounds investigated ylphenol

Injection Port Temperature



Care should be used when comparing quantitative results from methods with different extraction parameters

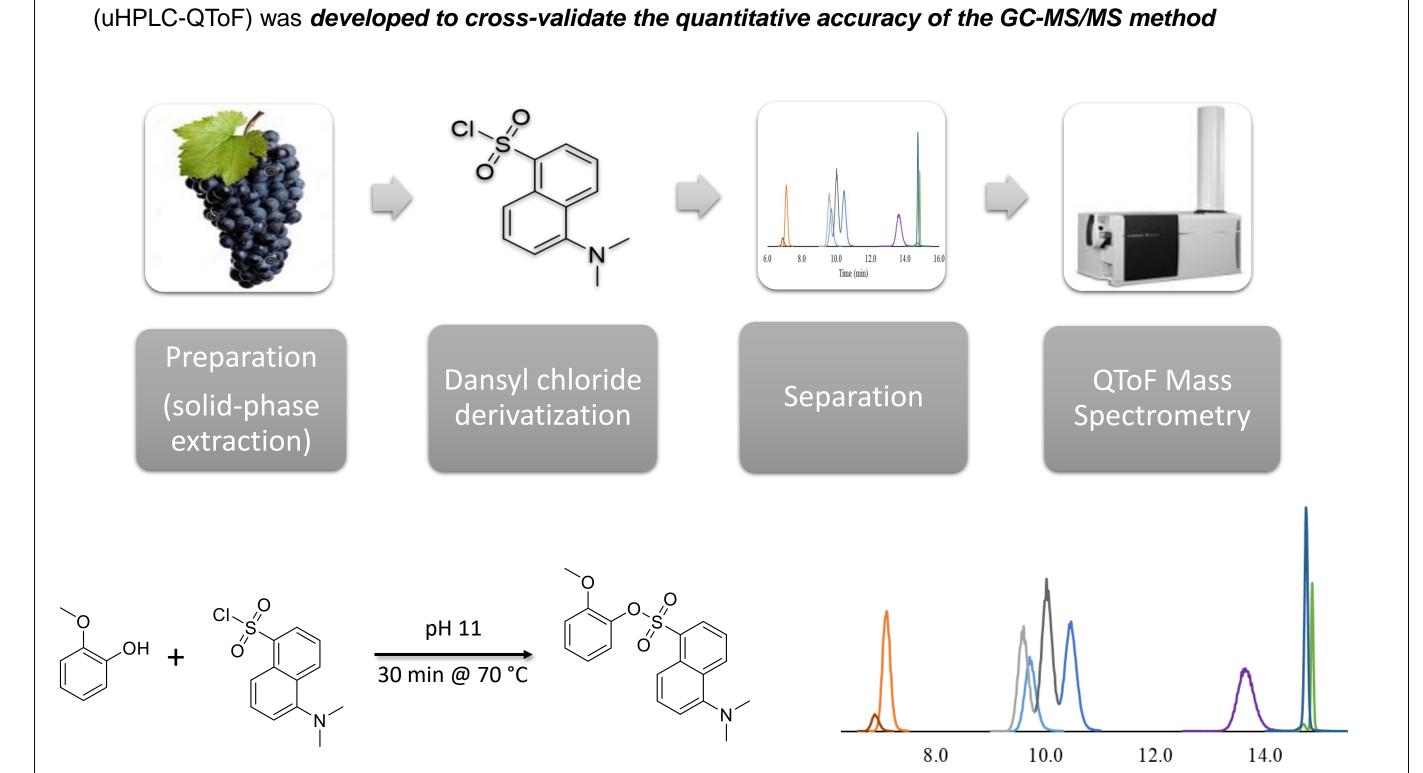
- Pollnitz, et al reported increased guaiacol response in wine as a function of injection port temperature ^g (top left)
- Attributed to thermal decomposition of co-extracted lignin
- Berry extracts show similar effect for guaiacol, but not d_3 -guaiacol
- Broad panel of volatile phenolics identified similar issues with eugenol and, more prominently, for syringol
- Existing methods use injection port temperatures that will yield inaccurately high concentrations for these compounds
- 220 °C selected as the optimal injection port temperature

Guaiacol

Dansyl chloride

Novel Analytical Method

Despite evaluation, injection port issue raised concerns regarding the quantitative accuracy of our GC-MS/MS method • Orthogonal method based on ultra-high pressure liquid chromatography-quadrupole time-of-flight mass spectrometry



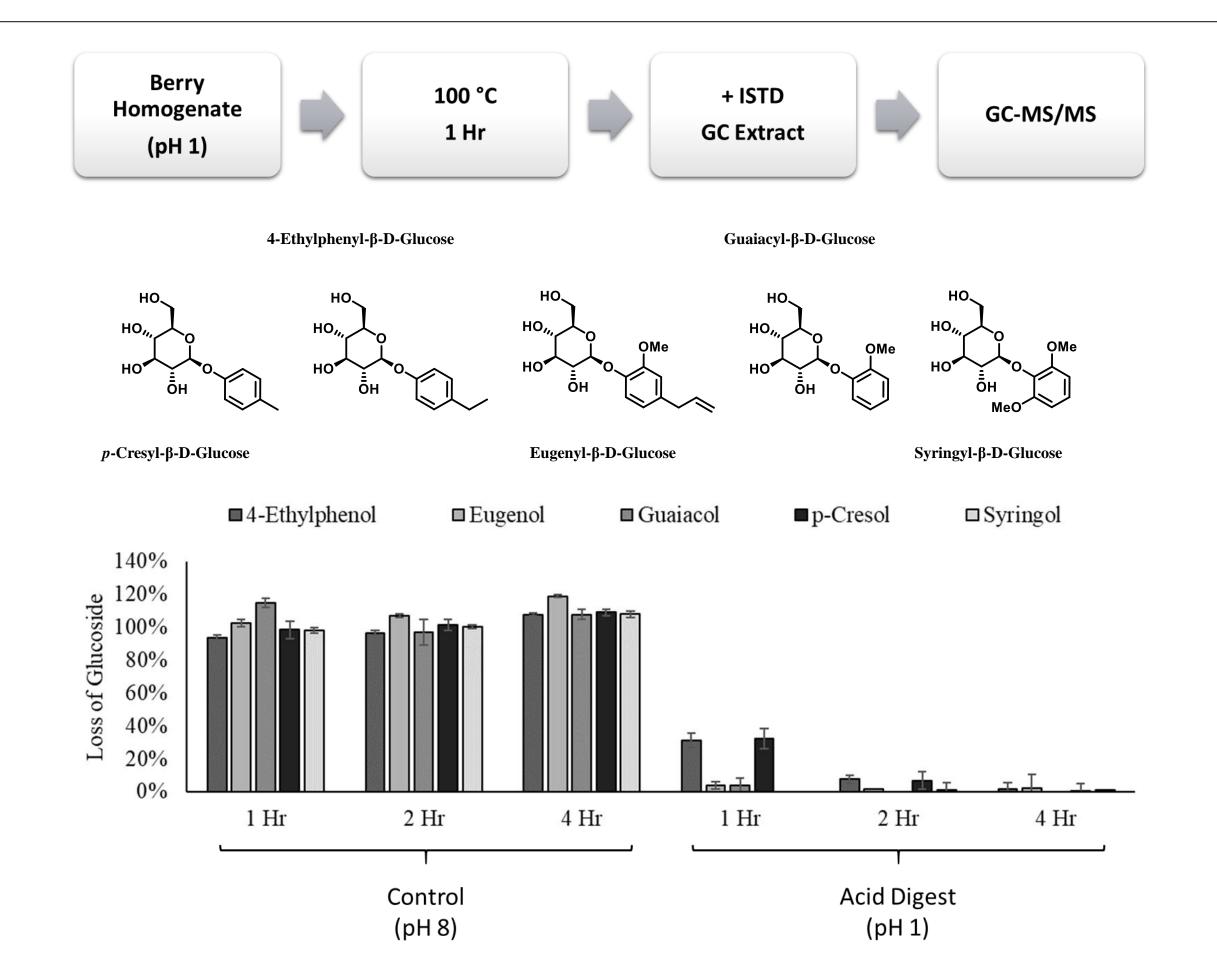
Time (min)

Method Validation

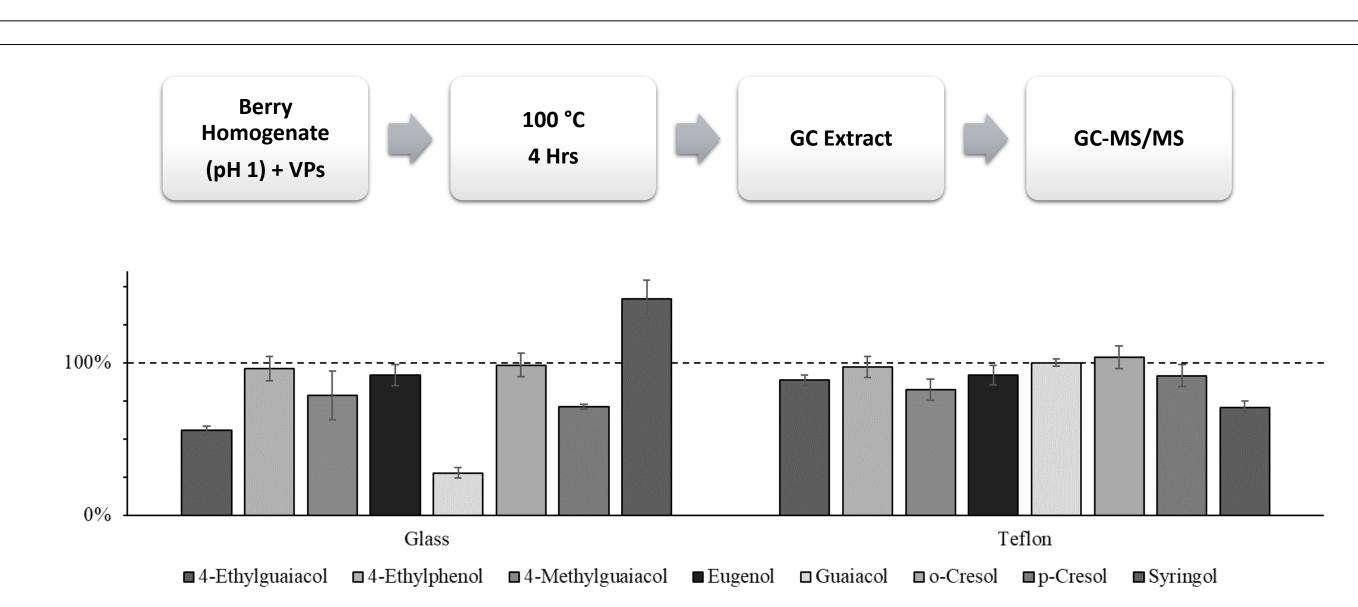
	GC-MS/MS			uHPLC-QToF				
Compound	LOQ (ng/g)	Accuracy (2/100 ng/g)	Repeatability (2/100 ng/g)	LOQ (ng/g)	Accuracy (5/200 ng/g)	Repeatability (5/200 ng/g)	HOR _{RAT}	
4-Ethylguaiacol	1.62	97/85	8/9	2.72	105/107	14/1	-	
4-Ethylphenol	0.55	95/82	5/5	3.39	90/100	17/2	-	
4-Methylguaiacol	0.49	90/75	6/3	1.22	98/111	9/4	-	
Eugenol	1.05	113/106	13/3	1.22	112/94	6/5	1.5	
Guaiacol	1.67	93/90	8/7	1.06	110/111	9/1	0.6	
<i>m</i> -Cresol	-			0.40	89/100	5/4	-	
o-Cresol	0.85	86/67	14/2	0.83	90/106	10/6	1.2	
<i>p</i> -Cresol	0.77	76/82	6/3	0.94	92/114	10/3	-	
Syringol	0.84	126/99	6/4	2.64	90/87	13/3	-	

- Both methods are suitably sensitive, precise and accurate for the analysis of smoke-exposed berries
- uHPLC-QToF method would be more sensitive if using a triple quadrupole detector
- Cross-validation using Horwitz ratios (HOR $_{RAT}$) demonstrated the quantitative equivalence of both methods and, by extension, the quantitative accuracy of the developed GC-MS/MS method (with optimized extraction and injection port parameters)

Acid Hydrolysis



- Model glucosides were synthesized to facilitate development of a method that yields quantitative recovery of free volatile phenolic compounds following acid-catalyzed hydrolysis
- Current methods use 1 Hr @ 100 °C, which is insufficient to hydrolyze the model glucosides (4 Hrs is required)



- Existing methods do not specify reaction vessels for acid digests
- Our data show that quantitative recovery of liberated volatile phenols is not possible in glass vials, while switching to Teflon vials lead to near quantitative yields
- Cause of this effect is unclear, although our data suggest it is a function of non-specific surface interactions under the harsh hydrolytic conditions employed

Summary and Future Works

- Predicting the presence of smoke-taint in wine based on the chemical composition of smoke-exposed berries will require a quantitatively accurate assessment of the total volatile phenol load in the berry
- Existing methods should be carefully evaluated regarding their extraction parameters and, in the case of GC-MS/MS, their injection port temperatures
- Independent and cross-validation of the developed GC-MS/MS method demonstrated quantitative accuracy
- Acid-catalyzed hydrolysis required 4 hours to liberate bound glycosides and needed to be carried out in teflon vials to ensure quantitative recovery of volatile phenolic compounds
- The developed quantitative work-flows will be applied to the analysis of field trials from 2016
- Build model for predicting sensory properties of wine based on VP composition

Acknowledgements

Brandon Whitmore (glucosides) University Graduate Fellowship Walter C Sumner Memorial Fellowship





References

a) Parker, et al. 2012. Aust. J. Grape Wine Res. 21, 537. b) Hayasaka, et al. 2010, J. Ag. Food Chem. 58(20), 10989. c) Wilkinson et al, Aust. J. Grape Wine Res. 2011, 17, 22. d) Pollnitz et al, J. Agric. Food Chem. 2004, 52, 3244. e) Singh et al, Aust. J. Grape Wine Res. 2011, 17, 13. f) Singh et al, Curr. Bioact. Comp., 2012, 8, 190. g) Pollnitz et al, J. Agric. Food Chem. 2004, 52, 3244.

